249. Piperaceae Alkaloids: Part IV¹) Structure and Synthesis of Cyclostachine A, Cyclostachine B and Cyclopiperstachine²)

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Dedicated to Professor T. R. Govindachari on the occasion of his 60th birthday

Summary. Three new alkaloids designated as cyclostachine A (2), cyclostachine B (7) and cyclopiperstachine (10) have been isolated from *Piper trichostachyon C. DC*. Their structures have been derived on the basis of spectral and degradative studies and confirmed by synthesis. The ¹H- and ¹³C-NMR. spectra of these compounds are discussed.

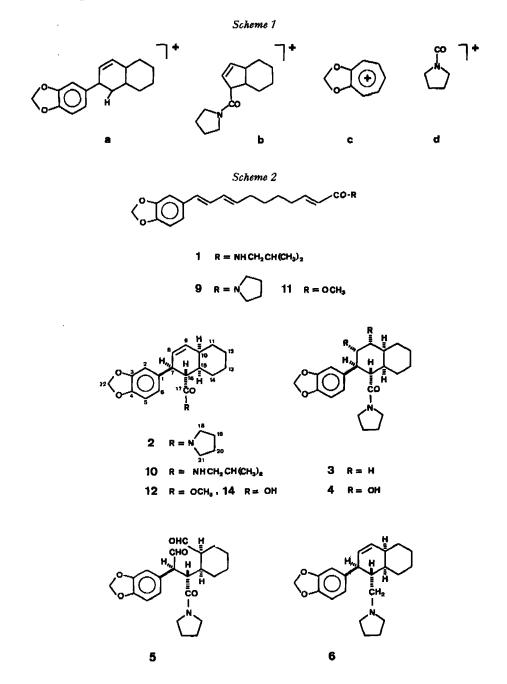
The isolation and structure elucidation of the alkaloid piperstachine (1) (from *Piper trichostachyon C.DC.*) were reported recently [3]. This structure was confirmed subsequently by synthesis [1]. Besides piperstachine, we have isolated three closely related alkaloids – cyclostachine A (2), cyclostachine B (7) and cyclopiperstachine (10). All three alkaloids are racemic as shown by CD. and ORD. determinations. We report here the structure elucidation and synthesis of these alkaloids. A preliminary communication on the structure of cyclostachine A has already been published [4]. The alkaloids, isolated by cold percolation of the plant with hexane, were purified by chromatography.

Cyclostachine A (2). - Cyclostachine A, m. p. 136–138°, $C_{22}H_{27}NO_3$ (M⁺: m/e 353), has in the IR. spectrum ν_{max}^{KBr} 1630 cm⁻¹ (tertiary amide). Its ¹H-NMR. spectrum showed the presence of a methylenedioxyphenyl group, an isolated disubstituted double bond and an acylpyrrolidine moiety. The ¹H-NMR. spectral assignments are presented in Table 1. The ¹³C-NMR. spectrum of cyclostachine A showed the absence of quaternary carbon atoms in the aliphatic region. (For the ¹³C-NMR. data see Table 2 and discussion on page 2300). The UV. spectrum, λ_{max}^{EtoH} 235, 287 nm (log e 3.65, 3.62), showed the absence of conjugation of the double bond with the aromatic ring. Its mass spectrum exhibited the molecular ion peak at m/e 353 and prominent ions at m/e 255, 218, 135 and 98 due to the ions (a) – (d), the ions (b) and (c) arising by rearrangement.

¹⁾ For Part III, see [1].

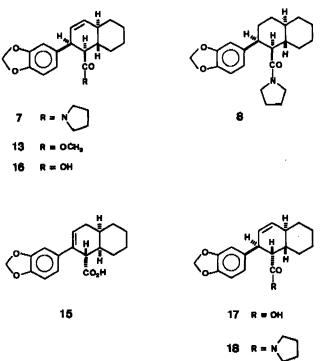
³) Contribution No. 410 from *Ciba-Geigy Research Centre*; ¹³C-NMR. Spectroscopy, Part 13. For Part 12 see [2].

Acid hydrolysis of cyclostachine A gave pyrrolidine identified by GC. and MS. [5]. Catalytic reduction afforded the dihydro derivative 3. The presence of a disubstituted double bond was confirmed by OsO₄-Oxidation to the diol 4 which on cleavage with periodate gave the dialdehyde 5. The ¹H-NMR. spectrum of the latter showed the presence of two aldehyde protons – one as a doublet at 9.67 ppm (J =





Scheme 3



2 Hz) and the other as an apparent triplet at 9.74 ppm ($\overline{J} = 1.5$ Hz) wherein the additional splitting is probably due to long-range coupling. Reduction of cyclostachine A with lithium aluminium hydride gave the amine 6 isolated as its sulfate, m. p. 110°, the structure of which was determined by X-ray crystal analysis [4]. Fig. 1 depicting molecule 6 as found in the crystal shows that atoms C(7), C(8), C(9), C(10) and C(15) in the cyclohexene ring are coplanar and C(1), C(14) and C(17) are all axially disposed. This is in contrast to ¹H-NMR. results which show that the amide 2 in solution has these three substituent atoms equatorial (see formula below). The nearly *trans*-diaxial arrangement of H(15), H(16) and H(7) is in good agreement with the observed coup-

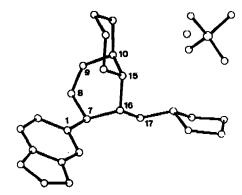
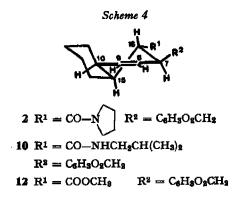


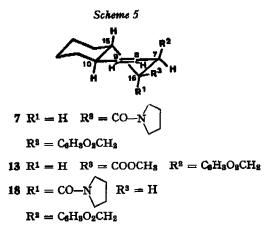
Fig. 1. Stereoscopic drawing of the structure of 6-hydrogensulfate [4]

ling constants ($J_{15,16} = 11.0$ Hz, $J_{7,16} = 10.0$ Hz). The pseudo-axial position of H(7) is confirmed by the rather large allylic coupling (2.5 Hz) with H(9). The H(9)–C(9)–C(8) plane and the projection of the C(7)–H(7) bond form an angle of ~80°, and maximum allylic coupling ${}^{4}J_{H,H}^{*}$ is expected for 90°. Also the 5 Hz coupling of H(9)



with H(10) is consistent with a torsional angle of $\sim 30^{\circ}$ (measured in *Dreiding* model). The experimentally observed large vicinal coupling constants between H(7), H(16) and H(15) cannot be explained on the basis of the conformation shown in Fig. 1 or by boat conformations of the cyclohexene ring. The conformation of the amine in the solid state is different probably because of the greater volume required by the non-planar amine ring compared with the amide form and the need to proximate the sulfate ion to the nitrogen atom for the ionic and hydrogen-bonded linkage.

Cyclostachine B (7). – Cyclostachine B, m. p. 135–136°, $C_{22}H_{27}NO_3$, is isomeric with cyclostachine A and has similar UV., IR. and mass spectral behaviour (see Exper. Part) indicating it to be a stereoisomer of cyclostachine A. The assignments in its ¹H-NMR. spectrum are presented in Table 1, the ¹³C-NMR. data in Table 2. Catalytic reduction gave the dihydroderivative 8. The aryl-octahydronaphthalene skeleton of cyclostachine A (2) is clearly originating from a triene-amide 9 related to piperstachine (1). Intramolecular *Diels-Alder* reaction of 9 would be expected to give the two racemates 2 and 7. This suggested that cyclostachine B could have the



trans-stereochemistry depicted in formula 7. Its ¹H-NMR. spectrum is in excellent agreement with the conformation shown below. H(16) appears as a double doublet with $J_{16,15} = 10.5$ Hz and $J_{16,7} = 6.0$ Hz demonstrating an ax,ax arrangement of H(15) and H(16) and a pseudo-equatorial position of H(7). The axial position of H(16) is further supported by the chemical shift (2.80 ppm) which is very nearly the same as in cyclostachine A (2.76 ppm). The $J_{9,10}$ coupling is very small and not resolved in agreement with the torsional angle of ~ 80° in the above conformation. The splitting patterns of the two olefinic protons are compatible with the above structure, *i.e.* only slight broadening for the H(9) doublet and extensive splitting of the H(8) doublet due to vicinal coupling with H(7) and allylic coupling with H(10). Thus, the splitting patterns of H(8) and H(9) are just opposite to cyclostachine A, in agreement with the opposite conformation of the cyclohexene ring.

Cyclopiperstachine (10). – Cyclopiperstachine, m. p. 220°, $C_{22}H_{29}NO_3$, is isomeric with piperstachine (1) and has spectral properties (see Exper. Part) similar to cyclostachines A and B. Its MS. and ¹H-NMR. spectra show the presence of an isobutylamide group in place of the pyrrolidine moiety present in cyclostachines A and B. The mass spectrum exhibits the molecular ion peak at m/e 355 and prominent ions **a** and **c** at m/e 255 and 135. The ¹H-NMR. spectral assignments are shown in Table 1. These data suggest structure 10 for the alkaloid, the conformation shown above (as for 2) best explaining the H,H coupling constants between H(8), H(9), H(7) and H(10).

Synthesis. – Intramolecular *Diels-Alder* reaction of the ester 11 [1] gave a mixture of the two esters 12 and 13 which were separated by chromatography and crystallisation. The ¹H-NMR. spectrum of the major product supports its formulation as 12 with the conformation shown above for 2, in agreement with expectations [6][7]. The ¹H-NMR. spectrum of the minor isomer shows it to have the other possible structure 13 with the conformation given for 7.

Alkaline hydrolysis of ester 12 yielded the acid 14 which was converted to the amide 2, identical in all respects with naturally occurring cyclostachine A. Esterification of the acid 14 with diazomethane gave back the ester 12, thus proving the absence of any isomerisation during the alkaline hydrolysis. When vigorous conditions were employed in the hydrolysis, isomerisation took place to yield the acid 15, whose UV. spectrum showed the presence of a methylenedioxystyrene chromophore. Its ¹H-NMR. spectrum showed the presence of only one vinylic proton at 6.07 ppm (t, J = 3 Hz). Treatment of the acid chloride of 14 with isobutylamine gave the amide 10, identical in all respects with natural cyclopiperstachine. Amide 10 was also obtained by thermal cyclisation of piperstachine (1).

Alkaline hydrolysis of the ester 13 gave the acid 16 which was converted to the amide 7, identical in all respects with natural cyclostachine B. Vigorous conditions during the hydrolysis of 13 resulted in epimerisation at C(16) and yielded the acid 17. Whereas H(16) in acid 16 appears at 2.75 ppm as a double doublet, in acid 17 it occurs as a broad singlet at 2.6 ppm. Acid 17 was converted to the amide 18 which is an isomer of cyclostachines A and B. Its ¹H-NMR. spectrum is reported in Table 1 and agrees well with the conformation depicted for 7. In this compound, H(16) appears as a doublet (J = 3.5 Hz) at 2.72 ppm. Double resonance shows that the

Table 1. ¹ H-1 Compound	VMR. data H(2,5,6)	⊧ ^a) H(7)	H(8)	H(9)	H(10)
Cyclostachine A 2	6.7 m	3.68 d,q J _{7,16} 10.0	5.55 br d J _{8,9} 10	5,9 d,d,d	3.15 d,t ^J 10,11a ¹ ^J 10.11e ⁶
Cyclostachine B 7	6.4-6.7 m	3.0-4.0	5.55 m	5.7 d J _{9,8} 10	3.0-4.0
Amide 18	6,74 m	3.3-3.7	5.6 d,d,d J8,9 10 ^J 8,7 4 J8,10 ²	5.8 d,t J _{9,8} 10 J _{9,10} 1.5 J _{9,7} 1.5	3.3-3.7
Cyclopiper- stachine 10	6.64 m	3.66 m	5.5 brd J _{8,9} 10	5.8 d,d,d ^J 9,8 ¹⁰ ^J 9,10 ⁵ ^J 9,7 ³	2.0 T
Ester 12	6.5-6.8 m	3.65 d,q J7,16 ¹⁰	5.5 br d J _{8,9} 10	5.8 d,d,d J9,8 10 J9,10 4.5 J9,10 2.5	2.C m
Ester 13	6.4-6.8 m	3.5-3.8 m	5.4-) m		

*) All spectra were taken in CDCl₃ at 100 MHz except for compound 13 which was run at 60 MHz. Chemical shifts are in δ [ppm], s singlet, d doublet, t triplet, q quartet, m multiplet, br broad; J values are in Hz.

doublet splitting is due to coupling with H(15) which is at ~ 1.6 ppm, but not with H(7). Since the two substituents at C(16) and C(7) are both axial a minor distortion of the cyclohexene half chair to release steric 1,3-diaxial interactions with H(10) and H(15) respectively, would make the torsional angle for the H(7) and H(16) protons $\sim 90^{\circ}$ so that no coupling should be observed for these protons, as it is found in the spectrum.

The 13 C-NMR. spectral data (Table 2) fully support the structures of cyclostachine A (2), cyclostachine B (7) and of the synthetic stereoisomer 18. The spectra exhibit 22 lines in the case of 2 and 21 lines with one alightatic CH₂ signal of double

H(15)	H(16)	H(18)	H(21)	H(22)	Mísc.
2.0-2.5 m	2.76 d,d ^J 16,15 ^{11,0} ^J 16,7 ^{10,0}	3.4 t 0 J 6.5 0	2.0~2.5 m	5.90 8	
1.0-2.2	2,80 d,d ^J 16,15 ^{10,1} ^J 16,7 ^{6,0}	5		5.88 s	
1.6 m	2.72 d J _{16,15} 3.5	·		5.92 s	
2.4	2.3 m	2.9 t J _{18,19} 7 J _{18,NH} 7	0.68 0.72 d J _{20,19} 7 J _{21,19} 7 (CH- <u>Me</u> ₂)	5.86 \$	5.25 br t (NH) J _{NH,18} 7
2.4	2.75 d,d ^J 16,15 12 ^J 16,7 10			5.88 s	3.52 s (CO ₂ Me)
	2.75 d,d ^J 16,15 11 ^J 16,7 7			5,9 \$	3.42 s (CO ₂ Me)

intensity in the cases of 7 and 18. Off-resonance decoupling experiments reveal the presence of four singlets (C(1), C(3), C(4) and C(17)), five doublets in the olefinicaromatic region (C(2), C(5), C(6), C(8) and C(9)), four doublets in the aliphatic region (C(7), C(10), C(15) and C(16)) and nine triplets (C(11), C(12), C(13), C(14), C(18), C(19), C(20), C(21) and C(22)). There are no signals of quaternary carbon atoms in the aliphatic region. Assignments of signals within these groups are given in Table 2. The spectral pattern in the low-frequency region (20-40 ppm) is very similar for 7 and 18 which both have a *trans*-octahydronaphthalene system, but differs considerably in 2 with a *cis*-skeleton. Further differences in the chemical shifts of the three stereoiso-

-	C(17)	C(3,4)	C(1)	C(6,8,9)	C(2,5)	C(22)	C(7,16)	C(18,21)	C(10,15)	C(11,12 C(19,20	
	173,6	147.7	138.7	133.4	108.4	101.0	47.2	46.4	36.6	30.6	26.0
2		146.3		128.5	108.1		46.8	45.4	35.6	28.8	24.3
				121.0						26.4	22.1
	170.2	147.3	135.1	133.3	110.0	100.9	49.9	46.4	36.6	42.2	26.4(2)
7		146.5		127.5	107.7		42.6	45.5	30.7	33.3	24.3
				122.4						26.9	
	173.0	147.5	139.7	134.1	108.7	100,8	49.4	47.4	36.6	38.0	26.4(2)
18		145.8		125.4	108,1		44.0	45.6	30.6 ^{b)}	33.3	24.4
				121.0						26.9	

Table 2. 18C-NMR. data) of the cyclostachines and the synthetic amide 18

•) δ [ppm], ± 0.2 ppm.

b) Tentative assignment, doublet-structure not proven.

mers are observed, as expected, in the resonances of C(15), C(16) and C(7). In all three compounds the diastereotopic carbons C(18) and C(21) exhibit different resonance frequencies.

Cyclostachines A and B evidently arise by intramolecular cyclisation of the trieneamide 9 and cyclopiperstachine from piperstachine (1). These three compounds have been isolated from the *cold* hexane extract and evaporation of the extract *in vacuo*. Since the thermal cyclisation of piperstachine (1) seems to require a temperature of 110° , the three new compounds reported here do not appear to be artefacts formed in the isolation stage.

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Experimental Part

UV. and IR. spectra were determined on *Beckman* model DK-2A and *Perkin-Elmsr* model 421 spectrophotometers. MS. were recorded on an *Atlas Varian* Mat CH-7 spectrometer using direct inlet system. ¹³C-NMR. spectra were measured on an XL-100-15 spectrometer at 25.2 MHz in the pulsed mode, ¹H-NMR. spectra on a *Varian* A-60 instrument, including double resonance experiments on an HA-100 instrument. Deuteriochloroform as solvent and tetramethylsilane as an internal reference were used for ¹³C-NMR. spectra. Chemical shifts are given in ô-values (ppm), coupling constants in Hz.

Isolation. The powdered stem of Piper trichostachyon (6 kg) was extracted with hexane $(3 \times 30 \text{ l})$ by cold percolation. The extract was evaporated in vacuo to yield a greenish oil (80 g). This was dissolved in hexane and chromatographed over a column of neutral alumina (2 kg) and gradient-eluted with hexane, hexane/C₆H₆, C₆H₆/CHCl₃ and CHCl₂/MeOH. Fractions (300 ml) were collected and monitored by TLC. Fractions 16-20 (eluant: hexane/C₆H₆ 1:3) gave a yellow oil (15 g) which was rechromatographed over silica gel in C₆H₆ to give cyclopiperstachine (10) (70 mg), colourless needles (from CH₂Cl₂/hexane), m.p. 220°, TLC., silica gel, CHCl₂/MeOH 99:1.

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R_f 0.65. – UV. (λ_{max}^{EtOH}) : 236, 286 nm (log ε 3.67, 3.62). – IR. (ν_{max}^{KBr}) : 3340, 1645, 1535, 1505, 1490, 1470, 1445, 1390, 1370, 1360, 1350, 1300, 1260, 1215, 1200, 1190, 1160, 1105, 1045, 930, 880, 835, 805 cm⁻¹. – ¹H-NMR. see Table 1. – MS. (m/ε) : 355 $(M^+, 50\%)$, 272 (12), 256 (38), 255 (68), 254 (57), 240 (10), 220 (22), 162 (20), 148 (34), 141 (15), 135 (100), 121 (50), 115 (25).

C22H29NO9 (355,5) Calc. C 74.3 H 8.2 N 3.9% Found C 74.2 H 8.4 N 4.2%

Fractions 28-35 (eluant: C₆H₆) yielded colourless cubes of cyclostachine A (2) (1.2 g), m.p. 136-138° (from ether/hexane), TLC., silica gel, CHCl₈/MeOH 99:1, R_f 0.5. – UV. (λ_{max}^{EtOH}): 235, 287 nm (log e 3.65, 3.62). – IR. (γ_{max}^{KBR}): 1640, 1500, 1480, 1430, 1380, 1245, 1190, 1115, 1100, 1035, 970, 925, 900, 880, 870, 832, 812 800, 758, 750, 730, 720 cm⁻¹. – ¹H-NMR. see Table 1. – MS. (m/e): 353 (M^+ , 100%), 270 (45), 255 (44), 254 (90), 240 (18), 218 (45), 212 (8), 211 (15), 204 (8), 181 (6), 169 (6), 153 (10), 152 (10), 148 (12), 141 (14), 135 (78), 128 (20), 115 (25), 103 (12), 98 (95), 91 (24), 77 (22), 70 (24), 56 (35), 55 (92).

C22H27NO3 (353.5) Calc. C 74.8 H 7.7 N 4.0% Found C 74.7 H 7.9 N 3.8%

Fractions 39–41 (eluant: C₆H₆) yielded cyclostachine B (7) (250 mg), colourless needles (from CH₂Cl₂/hexane), m.p. 135–136°. – TLC., silica gel, CHCl₂/MeOH 99:1, R_f 0.6. – UV. (λ_{max}^{EtOH}): 236, 287 nm (log ε 3.73, 3.62). – IR. (ν_{max}^{KBx}): 1640, 1500, 1490, 1480, 1370, 1295, 1275, 1240, 1210, 1190, 1165, 1120, 1100, 1090, 1070, 1045, 995, 965, 940, 900, 890, 880, 828, 810, 790, 785, 735, 725, 700 cm⁻¹, – ¹H-NMR. see Table 1. – MS. (m/e): 353 (M⁺, 86%), 270 (8), 255 (35), 254 (21), 240 (14), 218 (100), 192 (11), 161 (13), 152 (28), 139 (22), 135 (40), 98 (35), 91 (21), 70 (30), 55 (35).

C22H27NO8 (353.5) Calc. C 74.8 H 7.7 N 4.0% Found C 74.5 H 8.0 N 4.0%

Hydrolysis of cyclostachine A. Cyclostachine A (2) (100 mg) was heated with $6 \times HCl$ (4.5 ml) in a sealed tube at 100–110° for 16 h. The solution was extracted with CH_2Cl_3 to remove nonbasic impurities. The aqueous acid solution was evaporated *in vacuo*, the residue treated with a few drops of methanolic NH_3 and the solution used as such for GC. Part of the solution was adsorbed on chromosorb for mass spectrum. The product was identical with authentic pyrrolidine in retention time in GC. (Instrument: Varian VA 2740) column chromosorb 103, length of column: 6', internal diameter 3 mm, hydrogen flow rate 40 ml/min, column temp. 159°, injection block temp. 270°, detector temp. 300°, detector: FID).

Its MS., m/e 71 (M^+ , 14%), 70 (18), 43 (82), 42 (12), 28 (100), was identical with the reported MS. of pyrrolidine [5].

Dihydrocyclostachine A (3). A solution of cyclostachine A (100 mg) in EtOH (15 ml) was shaken with H₂ at atmospheric pressure over Pd/C catalyst (5%, 100 mg). The solution was filtered, evaporated and the gum crystallised from ether/hexane to yield dihydrocyclostachine A (40 mg), m.p. 165°. – UV. ($\lambda_{max}^{\text{EtOH}}$): 233, 287 nm (log ε 3.66, 3.58). – MS. (m/e): 355 (M⁺, 58%), 272 (20), 257 (32), 256 (95), 247 (16), 246 (23), 242 (15), 220 (28), 175 (15), 162 (20), 148 (44), 135 (100), 131 (18), 113 (60), 103 (17), 98 (60), 70 (35).

C12H H199NO8 (355.5) Calc. C 74.3 H 8.2 N 3.9% Found C 74.5 H 8.5 N 4.3%

Diol 4. A solution of cyclostachine A (190 mg) in dioxane (5 ml) was treated with OsO_4 (200 mg) and pyridine (0.2 ml) and left for 40 h at room temp. The complex was decomposed with H₂S, the solution filtered and the filtrate evaporated *in vacuo*. Crystallisation of the residue from EtOAc/hexane yielded the diol 4 (140 mg), m.p. 167°. - ¹H-NMR. (CDCl₃): 6,75 (3 H, m); 5.88 (2 H, s). - MS. (m/s): 369 (M⁺-H₂O, 100%), 340 (25), 286 (25), 270 (85), 253 (40), 246 (60), 193 (74), 175 (40), 135 (75), 98 (52).

C22H29NO5 (387.5) Calc. C 68.2 H 7.5% Found C 68.5 H 7.7%

Dialdshyde 5. A solution of the above diol (250 mg) in MeOH (10 ml) was treated with a solution of sodium metaperiodate (300 mg) in H₂O) in H₃O (10 ml) and allowed to stand at room temp. for 20 h. Dilution with H₂O, extraction with CHCl₃ and chromatography of the product over silica gel in CHCl₃/EtOAc 1:1 gave the dialdehyde 5 (55 mg), m.p. 116° (from EtOAc/hexane). – IR. (r_{max}^{KBr}): 2830, 2700, 1710 cm⁻¹. – ¹H-NMR. (CDCl₃): 9.74 (1H, t, $\overline{J} = 1.5$); 9.67 ppm (1H, d,

J = 2; 6.6-6.85 (3H, m). - ¹³C-NMR. (CDCl₃): 205.3, 199.2 (2 - CHO). - MS. (m/e): 385 (M+, 5%), 339(35), 274(35), 257(26), 246(74), 240(72), 204(63), 175(50), 135(54), 98(100).

C22H27NO5 (385.5) Calc. C 68.5 H 7.1% Found C 68.5 H 7.4%

LiAlH₄-reduction of cyclostachine A. A solution of cyclostachine A (500 mg) in dry ether (40 ml) was added with stirring to LiAlH₄ (500 mg) in ether (40 ml) under N₂. The mixture was stirred at 50° for 5 h, cooled, decomposed with 20% H₂SO₄ (50 ml) and left at room temp. for 2 days. The solid was filtered off, washed with ether and a little H₂O, and crystallized from McOH to yield the sulfate of amine 6 (100 mg), m.p. 110°. – UV. (λ_{max}^{EtOH}): 235, 287 nm (log ε 3.62, 3.58).

$$\begin{array}{cccc} C_{22}H_{29}NO_2 \cdot H_2SO_4 \cdot \frac{1}{2} H_2O & Calc. & C 59.2 & H 7.2 & N 3.1 & S 7.2\% \\ (446.6) & Found , , 59.3 & , 7.6 & , 3.3 & , 7.4\% \end{array}$$

The free amine, obtained as a gum, showed mass spectral peaks at m/s 339(M^+ , 10%), 268(8), 254(7), 225(7), 211(15), 198(7), 186(28), 181(8), 169(8), 165(10), 153(16), 152(16), 148(10), 141(24), 135(60), 131(16), 129(22), 128(32), 127(16), 115(42), 103(32), 91(42), 85(72), 84(100), 77(38), 70(36), 55(70).

Dihydrocyclostachine B (8). Cyclostachine B (150 mg) in EtOH (25 ml) was hydrogenated at atmospheric pressure over Pd/C catalyst (10%, 150 mg) to yield dihydrocyclostachine B (40 mg), m.p. 95° (from ether/hexane). – UV. (λ_{max}^{EtOH}): 233, 287 nm (log ε 3.69, 3.61). – MS. (*m/e*): 355 (*M*+, 30%), 260 (15), 256 (17), 247 (5), 246 (7), 242 (7), 220 (46), 161 (12), 148 (28), 135 (52), 131 (12), 113 (100), 98 (35), 70 (15).

C22H29NO3 (355.5) Calc. C 74.3 H 8.2 N 3.9% Found C 74.2 H 8.5 N 4.3%

Synthesis. – Cyclisation of methyl 11-(3, 4-methylenedioxyphenyl)-undeca-2, 8, 10-trans, trans, trans-trien-oate (11). A solution of the triene ester 11 (1.6 g) in xylenc (30 ml) was refluxed under N₂ for 3 h, evaporated in vacuo and the residue crystallized from other/hexane to yield the ester 12 (0.6 g), m.p. 136°. – UV. (λ_{max}^{EtOH}) : 235, 286 nm (log ε 3.60, 3.64). – IR. (ν_{max}^{KBr}) : 1720 cm⁻¹. –¹H-NMR. see Table 1. – MS. (m/e): 314 $(M^+, 54\%)$, 254 (100), 212 (18), 211 (24), 206 (22), 181 (40), 173 (44), 169 (36), 148 (90), 135 (95), 131 (90), 119 (85), 115 (72), 103 (45), 91 (82).

C19H22O4 (314.4) Calc. C72.6. H7.1% Found C72.4 H7.3%

Chromatography of the product from the mother liquor of the above ester over alumina in hexane yielded the isomer 13 (0.3 g), m.p. 87° (from ether/hexane). – UV. (λ_{max}^{EtOH}) : 237, 286 nm (log ε 3.65, 3.60). – IR. (ν_{max}^{KBr}) : 1725 cm⁻¹. – ¹H-NMR. see Table 1. – MS. (m/e): 314 $(M^+, 100\%)$, 255 (28), 254 (62), 240 (6), 232 (20), 211 (8), 206 (20), 187 (32), 173 (55), 157 (28), 148 (65), 135 (86).

C₁₉H₂₂O₄ (314.4) Calc. C 72.6 H 7.1% Found C 72.6 H 7.3%

Hydrolysis of the ester (12). - a) Acid 14. The ester 12 (0.3 g) in dioxane (8 ml) was refluxed with KOH (3 g) in MeOH (2 ml) and H₂O (2 ml) for 3 h, evaporated *in vacuo*, diluted with H₂O and extracted with ether. The aq. solution was acidified and extracted with ether to yield the acid 14 (250 mg), m.p. 140-143° (from ether/hexane). - UV. $(\lambda_{max}^{\text{EtOH}})$: 235, 286 nm (log ε 3.72, 3.65). - IR. (y_{max}^{KBr}) : 1702 cm⁻¹. - MS. (m/e): 300 (M⁺, 100%), 255 (25), 254 (24), 218 (16), 206 (24), 192 (32), 187 (20), 173 (46), 148 (85), 135 (60), 115 (30).

C₁₈H₂₀O₄ (300,3) Calc. C 72.0 H 6.7% Found C 72.0 H 7.0%

Re-esterification of the acid with ethereal diazomethane yielded the ester 12 identified by mixed m.p. and IR. spectrum.

b) Acid (15). The ester 12 (2 g) in 2-ethoxy-ethanol (30 ml) was refluxed with KOH (12 g) and H₈O (3 ml) for 45 min. The solution was evaporated *in vacuo*, the residue diluted with H₂O and worked-up as usual. The gummy acidic product was chromatographed over silica gel in hexane/ EtOAc 9:1. The initial fractions yielded the acid 14 (0.5 g). Later fractions gave the isomer 15 (0.2 g), m.p. 158° (from ether/hexane). – UV. (λ_{max}^{EtOH}) : 208, 257, 295 nm (log ε 4.40, 3.95, 3.72). – IR. (ν_{max}^{KBr}) : 1705 cm⁻¹. – ¹H-NMR. (CDCl₂ + DMSO-d₆): 10.33 (1H, s, OH); 6.7-6.9 (3 H, m);

6.07 (1 H, t, J = 3); 5.9 (2 H, s); 3.36 (1 H, br. s); 2.2 (4 H, br. s); 1.5 (8 H, br. s). – MS. (m/e): 300 (M^+ , 100%), 255 (28), 218 (15), 206 (30), 192 (28), 187 (21), 173 (39), 162 (20), 148 (52), 135 (45).

 $C_{18}H_{20}O_4$ (300.3) Calc. C 72.0 H 6.7% Found C 72.1 H 7.0%

Hydrolysis of the ester (13). – a) Acid (16). The ester 13 (100 mg) was refluxed mildly for 2 h with KOH (150 mg) in MeOH (2 ml) and H₂O (0.2 ml) to yield the acid 16 as a gum. – UV. (λ_{max}^{EtOH}): 236, 286 nm (log ε 3.69, 3.64). – ¹H-NMR. (CDCl₃): 10.51 (1 H, s, OH); 6.7 (3 H, br. s); 5.92 (2 H, s); 5.65 (2 H, br. s); 3.75 (1 H, br. s); 2.75 (1 H, d×d, J = 8 and 10).

b) Acid (17). The ester 13 (1 g) in dioxane (5 ml) was refluxed for 4 h with KOH (3 g), MeOH (5 ml) and H₂O (3 ml). The crude acid obtained was chromatographed over silica in hexane/EtOAc 9:1 to yield the acid 17 (0.5 g), m.p. 180° (from CH₂Cl₂/hexane). – UV. (λ_{max}^{EtOH}) : 236, 286 nm (log ε 3.67, 3.65). – IR. (ν_{max}^{KBr}) : 1698 cm⁻¹. –1H-NMR. (CDCl₃): 11.1 (1 H, br. s, OH); 6.77 (3 H, s); 5.93 (2 H, s); 5.7 (2 H, br. s); 3.75 (1 H, br. s); 2.6 (1 H, br. s). – MS. (m/e): 300 (M⁺, 72%), 255 (16), 254 (15), 240 (6), 192 (54), 187 (10), 173 (16), 148 (100), 135 (35), 115 (15).

C₁₈H₂₀O₄ (300.3) Calc. C 72.0 H 6.7% Found C 72.1 H 7.0%

Cyclostachine A (2). The acid 14 (80 mg) in C_6H_6 (5 ml) was refluxed mildly with oxalyl chloride (0.3 ml) for 1 h. The solution was evaporated *in vacuo*, dried well, redissolved in C_6H_6 (5 ml) and treated with pyrrolidine (1 ml). The solution was left overnight at room temp., washed (dil. HCl, H₂O), dried (Na₂SO₄) and evaporated. Crystallization of the residue from ether/hexane yielded the amide 2, identical (mixed m.p., UV., IR., ¹H-NMR. and MS.) with cyclostachine A.

C22H27NO3 (353.5) Calc. C74.8 H7.7 N4.0% Found C74.9 H7.9 N4.0%

Cyclostachine B (7). The acid 16 (150 mg) in C_6H_6 (3 ml) was refluxed for 1/2 h with oxalyl chloride (0.5 ml). The acid chloride obtained was treated with excess pyrrolidine as above to yield, after chromatography over alumina in hexane/CH₂Cl₂ (8:2), the amide 7 (50 mg), m.p. 135–136° (from ether/hexane), identical (mixed m.p., UV., IR., ¹H-NMR. and MS.) with cyclostachine B.

A mide 18. The acid 17 (250 mg) in C_6H_6 (5 ml) was refluxed mildly for $1^{1}/_2$ h with oxalyl chloride (1 ml). The acid chloride obtained was condensed with pyrrolidine (1 ml) to yield the amide 18 (150 mg), m.p. 150° (from $CH_2Cl_2/hexane$). – UV. (λ^{EtOH}_{max}): 235, 286 nm (log ε 3.71, 3.65). IR. (ν_{max}^{KBr}): 1635 cm⁻¹. – ¹H-NMR. see Table 1. – MS. (m/e): 353 (M^+ , 86%), 270 (30), 255 (30), 254 (44), 246 (10), 240 (18), 218 (76), 135 (100).

C22H27NO3 (353.5) Calc. C 74.8 H 7.7 N 4.0% Found C 74.5 H 8.0 N 4.3%

Cyclopiperstachine (10). – a) The acid 14 (250 mg) in C_6H_6 (5 ml) was refluxed mildly with oxalyl chloride (1 ml) for 1 h and the acid chloride obtained was treated with isobutylamine (1 ml) to yield the amide 10 (180 mg), m.p. 220° (from $CH_2Cl_2/hexanc$) identical (mixed m.p., UV., IR., ¹H-NMR. and MS.) with cyclopiperstachine. b) A solution of piperstachine (1) (0.3 g) in xylene (10 ml) was refluxed for 3 h under N₂, evaporated *in vacuo* and the residue chromatographed over alumina in hexane/CH₂Cl₂ (9:1) to yield the amide 10 (0.1 g), m.p. 220°, identical (mixed m.p., IR. spectrum) with natural cyclopiperstachine.

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